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STRUCTURE OF COMPLESTATIN, A VERY STRONG INHIBITOR OF PROTEASE ACTIVITY OF COMPLEMENT IN THE HUMAN COMPLEMENT SYSTEM

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Summary: The structure of complestatin, which strongly inhibits the protease activity of complements in the human complement system, has been determined as shown in Fig. 2 mainly based on HMBC. Its structure is closely related to glycopeptide antibiotics.

Complestatin (I) is a peptide isolated from the mycelium of <u>Streptomyces</u> <u>lavendulae</u>¹⁾, and strongly inhibits the hemolysis of sensitized erythrocytes by the complement system²⁾. To the best of our knowledge, <u>I</u> is the most potent compound among the known inhibitors with anti-complement activity such as flufenamic acid³⁾, leupeptin⁴⁾ and K-76⁵⁾. In this paper we wish to report the structural determination of <u>I</u> facilitated by extensive use of Heteronuclear Multiple Bond Correlation (HMBC)⁶⁾.

The physicochemical properties of <u>I</u> were as follows: mp. >300°C (dec.), $[\alpha]_D^{24}$ =+24.5° (c=0.13, MeOH-0.01N NaOH 2:1); UV λ_{max}^{MeOH} (ϵ) 282 (13800) and 292 (13200) nm; IR ν_{max}^{KBr} 3400 (OH), 1650 and 1510 (amide) cm⁻¹; color reaction, positive to Liebermann and Ehrlich, negative to Molisch and ninhydrin; elemental analysis, found C 54.13, H 3.81, N 7.27, Cl 15.45%, calcd. for $C_{61}H_{45}O_{15}N_7Cl_6$, C 55.14, H 3.41, N 7.38, Cl 16.01; HR-FABMS; M⁺ (<u>m/z</u>), found 1325.1060, calcd. 1325.1110.

Due to severe overlapping of the 13 C signals in the aromatic region, the number of \underline{sp}^2 quaternary carbons was determined to be 24 by spin echo experiments⁷). The ¹H- and ¹³C-NMR spectral data summarized in Table 1 showed the presence of 1 X -NCH₃, 2 X CH₂, 6 X CH, 20 X -CH=, 24 X -C=, 1 X -COOH, 6 X - CO-NH- and 1 X -C=0.

Acid hydrolysis of <u>I</u> (1N-HCl/CH₃COOH, 105°C, 22 hr) gave three main products (<u>II</u>, <u>III</u> and <u>IV</u>). Two of them obtained in the ratio of 1:2 were identified as 4-hydroxyphenylglycine (<u>II</u>) and 3,5-dichloro-4-hydroxyphenylglycine (<u>III</u>) by spectral analysis. Their absolute configurations were determined to be both D by comparison with a standard compound ($[\alpha]_D^{25}$ =-128.7°; <u>II</u>, $[\alpha]_D^{25}$ =-125.5°), and an authentic sample with L-(+)-configuration

group	C-13	H-1 (J _{Hz})	group	C-13	H-1 (J _{Hz})	group	C-13	H-1 (J _{HZ})	
1 2 3 4 5 6 C=0 CH NH	127.8 128.2 115.3 157.1 115.3 128.2 171.3 55.8	7.109(8.0) 6.765(9.2) 6.765(9.2) 7.109(8.0) 5.065(7.2) 8.507(6.3)	1 2 3 4 D 5 6 C=0 CH CH2 NCH2	134.5 131.5 123.1 155.2 121.7 130.6 168.4 61.3 35.0 31.2	7.825(2.0,8.8) 6.867(2.6,8.2) 7.079(2.5,7.0) 7.192(2.0,8.6) 5.065(7.2) 3.050 2.985	9 C=0 CH CH2 NH 1 2 3 G 4 5	136.3 [°] 170.3 57.1 28.2 124.1 130.1 122.5 155.8 122.5	4.182 2.886, 3.5 8.863(6.8) 7.776	
1 2 3 4 B 5 6 C=0 CH NH	131.0 127.0 122.0 148.7 122.0 127.0 169.2 51.5	7.341 7.341 5.196(7.0) 8.741(6.2)	1 2 3 E 4 5 6 C=0 CH NH	126.4 129.5 131.1 139.4 149.6 110.5 167.6 55.0	5.108(3.5) 5.475(4.0) 5.563(8.4) 8.285(9.2)	6 αC=0 βC=0 obta: of e +87.1 IV9)	130.1 185.7 163.7 Lned b ndura 5°; <u>II</u> was	7.776 y acid hyd cidin ⁸⁾ (<u>I</u> , [α] ²⁵ =- determined	rolysis [α] ²⁵ = 81.6°). to be
1 2 3 4 C 5 6 C=0 CH NH	131.9 126.7 121.7 148.1 121.7 126.7 169.8 55.2	7.305 7.305 5.563(8.4) 7.877(7.8)	1 2 3 4 5 F 6 7 8	123.6 111.5 126.3 118.4 123.7 134.4 114.4	10.9 7.272(2.8) 7.435(9.0) 6.830(1.5,8.2) 7.249	2-(3 phen acid comp the car car	,5-dic) yl)-2,3 based arison re ex bon i bon (nloro-4-hyd 2-dihydroxy 1 on NMR sj with <u>III</u> . isted no n <u>I</u> , the C-2) of	acetic pectral Since ketal ketal IV was

Table 1 13 C and 1 H NMR spectral data of complexitatin (in DMSO-d₆)

present as a ketone in the parent compound¹⁰⁾.

Detailed analysis of the COSY spectrum of I revealed the presence of the following units; 4 X -CO-NH-CH-, 1 X -CO-NH-CH-CH₂-, 1 x -CH₂-CH-N-, 2 X 1,4disubstituted aromatic systems and two ortho-coupled aromatic protons.

Analysis of the HMBC spectrum of I established seven partial structures, i.e., six amino acid units and one fragment with a ketone function (A to F and G, respectively, in Fig. 1). The arrows in Fig. 1 indicate long range couplings between protons and carbons $({}^{2}J_{C-H} \text{ or } {}^{3}J_{C-H})$. It is clear that <u>II</u>, III. and IV obtained by acid hydrolysis were originated from unit A, units B and C, and unit G, respectively. Taking account of the overlapping carbonyl carbons, Fig. 1 accommodates all the carbons present in I.

Non equivalent proton chemical shifts of D-2 and D-6, and D-3 and D-5 of the symmetric aromatic side chain in partial structure D suggested that the oxygen atom at D-4 is protected by a bulky group which restricts free rotation of the aromatic ring D.

The large ${}^{1}J_{C-H}$ value 11 observed with F-2 proton together with positive Ehrlich and Liebermann reactions of I indicated the presence of an indole



ring in unit F. This structure is supported by the ${}^{2}J_{C-H}$ and ${}^{3}J_{C-H}$ long range couplings between F-2, F-5 and F-8 protons and relevant carbons shown in Fig. 1. 169.2 In addition, the direct linkage between F-7 and E-3 carbons was revealed by long range coupling between E-3 (δ_{C} 131.0) and the protons at F-6 ($\delta_{\rm H}$ 6.830) and F-8 ($\delta_{\rm H}$ 7.249), and F-7 (δ_{C} 134.5) and the protons at F-5 (δ_H 7.435) and E-2 ($\delta_{\rm H}$ 5.108).

The sequence of six amino acid units could be obtained by analysis of long range couplings between α -methine and/or amide protons and the carbonyl carbon of the adjacent amino acid fragment (see Fig. 1). The analysis was started from unit D, in which the long range couplings between the N-methyl protons and a carbonyl carbon ($\delta_{\rm C}$ 169.2) enabled to distinguish the carbonyl carbon ($\delta_{\rm C}$ 168.4) in the unit D from one belonging to the next amino acid residue ($\delta_{\rm C}$ 169.2). The latter carbonyl carbon is present in unit B connecting units B and D. By repeating this analytical procedure, the sequence A-D-B-E-C-F- could be established. Since unit G gave <u>IV</u> (vide supra), G must be located at the next position of unit F.



As explained above, the phenolic hydroxy group of unit D must be protected to restrict free rotation of the ring D. Since acid hydrolysis gave phenolic compounds <u>II</u>, <u>III</u> and <u>IV</u> (vide supra) which apparently derived from

units A, B, C and G, unit D must be combined to one of the hydroxy groups of unit E. This position was determined by observing the deuterium-induced upfield shift in the ¹³C-NMR spectrum of <u>I</u> taken in d₆-DMSO added with one drop of 1:1 mixture of H₂O and D₂O. The carbon signal of E-5 ($\delta_{\rm C}$ 149.6) showed no shift, while line broadening was observed with E-4 resonance ($\delta_{\rm C}$ 139.4) suggesting the linkage of unit D to E-5 carbon. Thus, the structure of complestatin is established as shown in Fig. 2.

<u>I</u> is structurally related to glycopeptide antibiotics such as vancomycin¹²⁾, ristocetin¹³⁾, teicoplanin¹⁴⁾ and chloropolysporin¹⁵⁾. The main differences are that <u>I</u> has no sugar units, and possesses the indole nucleus instead of the modified tyrosine or β -hydroxytyrosine unit.

It is interesting that the biological activities of \underline{I} and glycopeptide antibiotics are markedly different; the latter show very strong antibacterial activity to Gram positive bacteria, while \underline{I} inhibited the growth of a few Gram positive bacteria at a very high concentration (ca. 2000 µg/ml). Tested so far, glycopeptide antibiotics showed no anti-complementary activity. It is also very important that a deglycosylated derivative of chloropolysporin, which had almost same activity as the parent compound <u>in</u> <u>vitro</u>,¹⁶) was inactive against the complementary system¹⁷). These findings suggest that difference of the biological activities between glycopeptide antibiotics and complestatin is not due to the presence of sugar units in the former group.

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References and Footnotes

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ۆۋ	Physicochemical properties of IV were as follows: $C_{PH_4}O_{J_4}C_{J_2}$, HB-EIMS found 233.9503, calca. 233.9487; IR v (KBr) 1720, 1670, 1580 cm ⁻¹ ; NMR (in CD_30D) δ_{C_1} 171.7 (C-1), 101.5 (C-2), 131.1 (C-3), 128.1 (C-4, 8),
10)	123.3 (C-5, 7) and 151.3 (C-6), $\delta_{\rm H}$ 7.52 (singlet). Similar phenomenon was observed with phenylglyoxylic acid, which showed the molecular ion peak corresponding to the store form. On the other
	hand, the H-NMR spectrum of this compound indicated the presence of a 1:1 mixture of the ketone and hydrated forms 12 hours after dissolving in
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